THE PROTEOLYTIC ACTION OF THE SNAKE VENOM ENZYMES ARVIN AND REPTILASE ON N-TERMINAL CHAIN-FRAGMENTS OF HUMAN FIBRINGEN

Birgit HESSEL and Margareta BLOMBÄCK

Department of Blood Coagulation Research, Karolinska Institutet, Stockholm, Sweden

Received 9 August 1971

1. Introduction

Mammalian fibrinogens are built up by three peptide chains, $\alpha(A)$, $\beta(B)$ and γ [1]. The N-terminal ends of these chains are linked together in a firm "disulfide knot" (DSK) by means of disulfide bridges [2]. When thrombin acts on fibrinogen small fibrinopeptides are split off from the N-terminal ends of the $\alpha(A)$ and $\beta(B)$ chains. The result of this proteolysis is fibrin. The peptides, which have a molecular weight of less than 2000, are fibrinopeptide A and its analogues AP and AY from the $\alpha(A)$ chain and fibrinopeptide B from the $\beta(B)$ chain [3]. Fibrinopeptide AY has the same amino acid sequence as fibrinopeptide A minus the N-terminal alanine; AP is fibrinopeptide A with a phosphoserine residue replacing serine at position 3 from the N-terminal end. "A" refers to A+AP+AY. Polymerization starts when fibrinopeptide "A" is released. In its limited proteolytic action, thrombin rapidly hydrolyses the arginyl-glycine bonds binding the fibrinopeptides to the rest of the molecule. However, some other arginyl or lysyl bonds can also be split. In this category is the arginyl-valyl bond, occurring 3 residues from the arginyl-glyceryl bond split when the A-peptide is released [2, 4]. The tripeptide in question, Gly-Pro-Arg, (see fig. 1), has been isol-

Abbreviations:

DSK: Disulfide knot

A : Human fibrinopeptide A
AP : Human fibrinopeptide AP
AY : Human fibrinopeptide AY
B : Human fibrinopeptide B

"A": Human fibrinopeptide A+AP+AY

 $\alpha(\text{``A''})$ -chain fragment: Designates the N-terminal part of the $\alpha(\text{``A''})$ -chain fragment in the disulfide knot.

ated from thrombic digests of DSK and α "A"-chain-fragments of DSK [2, 4], as well as from plasmic digests of sulfitolyzed α (A) chain-fragment [5].

It has been shown that the thrombin-like enzyme, Reptilase, from the venom of *Bothrops atrox* has a more limited proteolytic action than thrombin. Only fibrinopeptides "A" were reported to be released from native fibrinogen [6, 7]. The same pattern of proteolysis has been reported for highly purified Arvin, another snake venom enzyme from *Agkistrodon rhodostoma* [8].

In our studies on the purified disulfide knot we found it of interest to compare the action of these two thrombin-like enzymes on this substrate. It was found that other bonds, in addition to the ones mentioned above, could be split by the enzymes.

2. Materials and methods

DSK was purified from cyanogen bromide (CNBr) treated human fibrinogen by gel filtration and counter current distribution [9]. Thrombin (EC 3.4.4.13) was prepared as earlier described [10]. The final preparation had an activity of 368 NIH units per mg. Reptilase R was obtained from Pentapharm AG, Basel. The final preparation had an activity equivalent to 95 NIH (thrombin) units/mg. Arvin was obtained in solution from Twyford Laboratories, London. The activity was found to be equivalent to 670 NIH (thrombin) units/ml. Digestions with the enzymes were performed at pH 6.5 and 8.5 for 2-4 hr at 37° [4]. The enzyme concentration was from 6-30 NIH units and the substrate concentration 5-10 mg per ml of final mixture. The digests were freeze-dried.

Low-voltage electrophoresis and fingerprint analysis on cellulose plates were performed as described earlier [4]. Gel filtration, chromatography on a Technicon Automatic Peptide Analyzer and amino acid analyses were performed as described [5, 11].

3. Results

Fig. 1 shows that all three enzymes released fibrino-peptides A and Ap from DSK and the α("A")-chain-fragment of DSK. Thrombin and Reptilase in addition released the Gly—Pro—Arg peptide which gives a yellow colour with ninhydrin. Arvin however, did not release this peptide. Instead another yellow peptide appeared. This peptide was isolated by preparative electrophoresis and by gel filtration and chromatography [5, 11]. Amino acid analysis gave the following residues: Glu 1.05, Pro 0.95, Gly 1.00, Val 1.77, Arg 1.98. The results suggested that a peptide containing the residues No. 17 to 23 of the α"A"-chain-fragment had been released by Arvin. Amino acid sequence with the manual PITC method of Edman [12] gave the expected sequence: Gly—Pro—Arg—Val—Val—Glu—Arg.

These results indicate that Arvin and Reptilase have different specifities.

The electrophoresis and fingerprint analysis also suggested that at least small amounts of B-peptide are indeed released by the venom enzymes when purified DSK and $\beta(B)$ -chain-fragment prepared from DSK are used as substrate. From Arvin and Reptilase digests of DSK not only B-peptide but also small amounts of a longer fragment was isolated by gel filtration and chromatography. This fragment, by amino acid analysis, was shown to be identical with the first 42 residues of the $\beta(B)$ -chain-fragment of the DSK [13]. Also other fragments are released, at least by Arvin, but not yet identified.

Acknowledgements

We wish to express our thanks to Dr. K. Stocker, Pentapharm AG, Basel, for the gift of Reptilase and for stimulating discussions. This work was supported by grants from NIH (HE 07379-07) and from the Swedish Medical Research Council (B 71-19X-520-07C and B 71-13X-2475-04C).

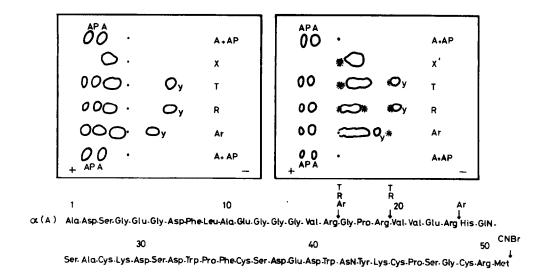


Fig. 1. Electrophoresis at pH 5.5 of thrombin, Reptilase and Arvin digests of α"A"-chain-fragment and DSK. Top left: substrate: α"A"-chain-fragment. Top right: substrate: DSK. X = undigested α"A"-chain-fragment; X¹ = undigested DSK. T = thrombin; R = Reptilase; Ar = Arvin. A = A-peptide [3]; AP = AP-peptide [3]. These were used as references. Spots were developed with ninhydrin spray, y = yellow spot. Below: Sequence of α"A"-chain-fragment of DSK from human fibrinogen [11].

References

- [1] B. Blombäck, Thromb. Diath. Hemorrhag. Suppl. 35 (1969) 161.
- [2] B. Blombäck, M. Blombäck, A. Henschen, B. Hessel, S. Iwanaga and K.R. Woods, Nature 218 (1968) 130.
- [3] B. Blombäck, M. Blombäck, P. Edman and B. Hessel, Biochim. Biophys. Acta 115 (1966) 371.
- [4] M. Blombäck, B. Blombäck, E.F. Mammen and A.S. Prasad, Nature 218 (1968) 134.
- [5] S. Iwanaga, P. Wallén, N.J. Gröndahl, A. Henschen and B. Blombäck, European J. Biochem. 8 (1969) 189.

- [6] B. Blombäck, Arkiv Kemi 12 (1958) 321.
- [7] B. Blombäck and M. Blombäck, Nouvelle Rev. Franc. d'Hématol. 10 (1970) 671.
- [8] M.R. Ewart, M.W.C. Hatton, J.M. Basford and K.S. Dodgson, Biochem. J. 118 (1970) 603.
- [9] B. Blombäck, B. Hessel and M. Blombäck, in preparation.
- [10] B. Blombäck and I. Yamashina, Arkiv Kemi 12 (1958) 299.
- [11] B. Blombäck, B. Hessel, S. Iwanaga, J. Reuterby and M. Blombäck, J. Biol. Chem., in press.
- [12] P. Edman, in: Protein Sequence Determination, ed. S.B. Needleman (Springer-Verlag, Berlin, Heidelberg, New York, 1970) p. 211.
- [13] B. Blombäck, M. Makino, S. Iwanaga, B. Hessel and M. Blombäck, in preparation.